An integrated approach for early in vitro seizure prediction utilising human-derived induced pluripotent stem cells and human ion channel assays **K L Rockley¹**; M Davis²; H Jennings¹; K Jones¹; P Levesque²; R A Roberts¹; M J Morton¹ ¹ApconiX, Alderley Park, Alderley Edge, Cheshire, UK; ² Bristol Myers Squibb, Princeton Pike, New Jersey, USA

Seizure liability remains a significant cause of attrition throughout drug candidates is not typically evaluated until the late stage of preclinical discovery, during in vivo toxicology studies. The timing of this assessment means that positive findings of seizure liability could result in the need to identify alternate clinical candidates. The resulting loss of competitiveness, delays, increased costs, and considerable safety risk all emphasize the need for improved methodologies to detect seizure liability earlier, ideally with reduced reliance on costly animal studies. High-throughput in vitro assays using human-derived induced pluripotent stem cell (hiPSC) neuronal cells coupled with screening seizure associated ion channels may offer an opportunity for a new paradigm in screening. A combined approach could provide mechanistic insight into off-targets causative of seizure and improve identification of potential seizure risks preclinically.

hipsc neuronal cell microelectrod

- iCell glutaneurons (FujiFilm CDI) containing 80% glutamatergic / 20%
- Electrical activity was monitored using the Axion Edge microelectrode
- The suitability of these cultures for seizure prediction was assessed by
- Comparisons to hippocampal slice data are also included for selected

EXAMPLE RASTER PLOTS



- The GABA receptor antagonists and pilocarpine show strong specificity for their targets

(MEA) instrument ation of seizurogen	 14 ion channels were selected based on Expression The activity of seizurogenic compounds was assess expressed in recombinant CHO or HEK cell lines Ion currents were measured by automated patch- 6 or 8-point curves were generated. An appropriate 												
unds (Fan et al., 20													
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ion ot with								CNS active therapi					
	Neanfime for Burst duration, hetwork burst duro hetwork burst duro network burst duro hetwork burst duro het									merion	romatine	Marami	ne ine
Amoxapine (3µM)	$\uparrow \uparrow$	NC	<u> </u>	$\downarrow \downarrow$	\checkmark				notar	i propiator	P ¹ 1 ² P ¹	onen vote	, ei
Bupropion (30µM)	\checkmark	\checkmark	<u>^</u> ^^	$\downarrow \downarrow$	$\downarrow \downarrow$	_ [¢]			Al.	Bri Qui	00 0	× 20.	<u> </u>
Chlorpromazine (3µM)	$\downarrow \downarrow$	$\downarrow \downarrow \downarrow$	<u> </u>	$\downarrow \downarrow$	$\downarrow \downarrow$	+ [¢]		Nav1.1					
Clozapine (3µM)	↓	↓	<u> </u>	$\downarrow \downarrow$	$\downarrow \downarrow$		_	Nav1.2					
Diphenhydramine (3µM)	↓	↓	<u> </u>	↓	$\downarrow \downarrow \downarrow$		_	Nav1.6					
Paroxetine (3µM)	<u>Т</u>	↓		<u>↓</u>	↓ ↓ ↓ ↓			Kv1.1					
Quetiapine (30µivi)	→ NC		NC					Kv2 1					
Enoxacin (10µM)	NC	NC	NC	NC	NC	∔ ¢	-	Kv2.1					
Pentylenetetazole (1mM)	¥	NC	NC	¥	NC	• + ^{↑φ}							
Picrotoxin (10µM)	, ↑	<u>ተተ</u>	1	↑	<u>^</u>	+ [↑]							
4-AP (100μM)	NC	\checkmark	NC	\checkmark	\downarrow	+ ^{†φ}		KV/.2//.3					
Linopirdine (10µM)	$\uparrow\uparrow$	NC	<u> </u>	\checkmark	^		1	KV/.3/7.5					
Pilocarpine (30uM)	\checkmark	NC	$\downarrow \uparrow$	^	^	<u>_</u> ¢		KCa1.1					
Strychnine (30µM)	1	NC	<u> </u>	\checkmark	\checkmark	+ ^{↑φ}		KCa4.1					
Acetaminophen (30µM)	NC	NC	NC	NC	NC	_ †		GABA α1β2γ2					
NC within 20% +,	个 /- 个个	2	0 - 50% iı 0 - 100%	ncrease increase	$\stackrel{\checkmark}{\downarrow} \stackrel{\checkmark}{\downarrow}$	20 - 50 50 - 10	0% decrease 00% decrease	Nicotinic α₄β₂ NMDA 1/2A					

Burst duration – Average time from the first spike to the last spike in a single electrode burst **Network burst frequency** – Total number of network bursts divided by the recording time **Network burst duration** - Average time from the first spike to last spike in a network burst Number of spikes per network burst - Average number of spikes in a network burst

DISCUSSION AND CONCLUSIONS

• The majority of seizurogenic compounds \uparrow network burst frequency, \downarrow burst/network burst duration and \downarrow the number of spikes per network burst • Exceptions include 4-AP and pilocarpine: 4-AP causes characteristic changes to the network burst pattern and pilocarpine decreases the frequency of network bursts • Of the GABA antagonists tested **picrotoxin** showed the most robust increase in activity in the MEA assay and inhibited the GABA $\alpha_1\beta_2\gamma_2$ ion channel • Amoxicillin and enoxacin showed no effects in the MEA assay and did not inhibit GABA $\alpha_1\beta_2\gamma_2$ in our ion channel assay • The Nicotinic $\alpha_{a}\beta_{2}$ channel was sensitive to the most compounds and the voltage-gated potassium channels were sensitive to more compounds than the sodium channels • The CNS active therapies inhibited the most ion channels outside of their MOAs, illustrating their promiscuity

• Our MEA data is largely concordant with findings in the rat hippocampal slice model, thereby illustrating the utility of the hiPSC neuronal cell MEA assay approach for early seizure prediction • These studies highlight the potential utility of hiPSC-neuronal assays and ion channel screening for early *in vitro* detection of seizure liability to support optimal drug design in early development



e risk hits (yellow):

Nicotinic $\alpha_{4}\beta_{2} = 21$, Kv2.1 = 15, Kv4.2 = 15, Kv7.3/7.5 = 14, Kv7.2/7.3 = 12, Kv1.1 = 10, GABA $\alpha_{1}\beta_{2}\gamma_{2} = 9$, Nav1.1 = 9, Nav1.2 = 8, Nav1.6 = 7, Kv3.1 = 6, KCa4.1 = 5, NMDA1/2A = 5, KCa1.1 = 0



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OR EARLY SEIZURE PREDICTION

profile, human mutations, function, pharmacology in the seizure related ion channels which were stably

mp (Q patch II, Sophion/ Patchliner, Nanion) ositive control was included for each ion channel

